



Transfection and induction of FLAG-HA tagged construct into cell culture:SC:Graveley

Revision as of 18:07, 13 June 2012 by [N.Washington](#) (Talk | contribs | block) (diff) ← Older revision | Current revision (diff) | Newer revision → (diff)

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Protocol Text

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FLAG-HA constructs were prepared according to [Construction of FLAG-HA expression clones:SC:Graveley](#) for a specific target. Plasmid DNA was then prepared using the PureLink™ HQ Mini Plasmid Purification Kit [Invitrogen K2100-01], and eluted with 50ul sterile water into a 1.5ml DNA LoBind microfuge tube (Eppendorf). The DNAs were quantified and assayed for A260/A280 ratio >1.8 using a NanoDrop spectrophotometer (ThermoFisher). Drosophila S2R+ cells were grown at 25°C in non-tissue culture treated polystyrene flasks (Corning Incorporated). Each clone was transiently transfected into a single 54ml culture of Drosophila S2R+ cells (1 x 10⁶ cells per ml) grown in a T-150 flask in Schneider's Drosophila Media (GIBCO 21720) with 10% heat-inactivated fetal bovine serum (GIBCO 26140079). Twelve micrograms of each DNA was combined with 300ul of Effectene (Qiagen) following the manufacturer's protocol. Twenty four hours after transfection, expression of the tagged protein was induced with 0.35mM CuSO₄. This level of CuSO₄ has been tested to induce a low-to-medium level of recombinant protein expression for a majority of representative clones. Cells were grown for 24 hours after induction.

Validation Form

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Protocol "Transfection and induction of FLAG-HA tagged construct into cell culture:SC:Graveley" (Version 4)

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Input type: ?

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Short Description: ?

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URL: ?

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