

# Differentiation of SK-N-SH Cells with *all-trans*-Retinoic Acid (RA) Prior to Harvesting

ENCODE4 - Version 1

## Materials List

1. MEM (CORNING Cellgro; Cat # 10-10-CV or 10-10-CM)
2. Heat Inactivated Fetal Bovine Serum (CORNING; Cat # 35-016-CV)
3. Penicillin-Streptomycin 10,000U/ml (Life Technologies, Cat # 15140 or Corning Cellgro, Cat # 300-002CI)
4. Phosphate Buffered Saline (1X PBS) w/o  $\text{Ca}^{2+}$ ,  $\text{Mg}^{2+}$  (CORNING Cellgro; Cat # 21-040-CM)
5. TrypLE Express (Life Technologies; Cat # 12604)
6. T75, T182 tissue culture treated flasks
7. Graduated pipets (2, 5, 10, 25, 50 ml)
8. DMSO (Fisher; Cat # BP-231-100)
9. Cryovials (Sarstedt; Cat # 72-694-006)
10. TC20 cell counter (Bio-Rad)
11. Counting Slides (Bio-Rad; Cat # 145-0011)
12. 0.40% Trypan Blue Dye (CORNING; Cat # 145-0013)
13. *all-trans*-Retinoic acid (Sigma, Cat # R2625)
14. Microscope

Note: Dissolve the *all-trans*-Retinoic acid in 100% of DMSO to make a 20 mM stock solution. Aliquot 100 or 150  $\mu\text{L}$  per vial. Keep in dark and stored in  $-20^{\circ}\text{C}$  freezer.

## Procedure

### Differentiation

Note: Volume (50 ml per flask) used in this protocol is for 182  $\text{cm}^2$  flasks.

- 1) Propagate cells until density reaches about 70% confluence.
- 2) Remove and discard culture medium.

- 3) Add 50 ml of growth medium containing all 15  $\mu$ L trans-retinoic acid stock solution (final concentration is 6  $\mu$ M).

Note: Turn off the light in the TC hood when adding the retinoic acid.

- 4) Cover the flask(s) with aluminum foil. Continue to culture for 48 hours.

### **Harvest**

- 1) Remove and discard culture medium.
- 2) Wash cells with room temperature (or warm) 1X PBS. (To remove all traces of serum that contains trypsin inhibitor.)
- 3) Add 10 ml (T182) of TrypLE and return to incubator for 7-15 minutes, or until cells detach.
- 4) 10-13 ml (T182) of complete growth medium and separate the cells by gentle pipetting.

Note: To avoid clumping DO NOT agitate the cells by hitting or shaking the flask while waiting for the cells to detach.

- 5) Examine viability and count cells using trypan blue staining (SOP).
- 6) Collect certain number of cells for your application.