

SOP: Propagation of IMR90 (ATCC CCL-186)

Information

Name: IMR90
ATCC #: CCL-186
Tissue: lung
Product Format: frozen
Morphology: fibroblast
Culture Properties: adherent
Biosafety Level: 1
Disease: normal: 16 weeks gestation Caucasian female

Lot number: 58607680 (ENCODE)

Materials List

1. MEM with 2mM L-glutamine and Earle's salts (Fisher Cat# 10-010-CM)
2. Heat Inactivated Fetal Bovine Serum (Life Technologies Cat# 10082147)
3. Non-essential Amino Acids, 100X solution (Life Technologies Cat# 11140-050)
4. Sodium Pyruvate, 100 mM (Fisher Cat# 25-000-CI)
5. Penicillin-Streptomycin 10,000U/mL (Life Technologies Cat#15140)
6. Phosphate Buffered Saline (1X PBS) (CORNING Cellgro, Cat# 21-040-CM)
7. TrypLE Express (Life Technologies Cat#12604)
8. T75, T525 culture flasks
9. Graduated pipets (1, 5, 10, 25, 50 mL)
10. Freezing medium (growth medium containing 95%; DMSO, 5%)
11. DMSO (Fisher Cat#BP-231-100)
12. Cryovials (Sarstedt Cat #72-694-006)
13. TC20 cell counter (Bio-Rad)
14. Counting Slides (Bio-Rad 145-0011)
15. Microscope

Growth Medium for IMR90

MEM with 2 mM L-glutamine and Earle's salts
10% FBS
Non-essential Amino Acids (1X)
Sodium Pyruvate (1 mM)
Pen-Strep (1X)

Procedure

A. Receipt of Frozen Cells and Starting Cell Culture

- 1) Immediately place frozen cells in liquid nitrogen freezer storage until ready to culture.

- 2) When ready to start cell culture, quickly thaw ampoule in a 37°C water bath.
- 3) As soon as ice crystals disappear, swab outside surface of the ampoule with 70% ethanol, then dispense contents of ampoule into a T75 flask with 20 ml of warm growth media.
- 4) Allow cells to recover overnight in 37°C, 5% CO₂ humidified incubator.
- 5) The next morning, the diluted DMSO-containing shipping/cryopreservation medium is aspirated from the cell layer and replaced with fresh medium.

B. Sub-culture

Volumes used in this protocol are for 75 cm² and/or 525 cm² flask; proportionally reduce or increase amount of dissociation medium for culture vessels of the other size.

- 1) Propagate cells until density reaches 70-80% confluence.
- 2) Aspirate medium.
- 3) Wash cells with room temperature (or warm) 1XPBS.
- 4) Add 3 mL (T-75) or 30 mL (T525) of TrypLE and return to incubator for 3 minutes, or until cells detach.
- 5) Add 7 ml (T-75) or 30 mL (T525) of complete medium and aspirate the cells by gentle pipetting.
- 7) Perform 1:2 to 1:8 cell split as needed.
- 8) Incubate at 37°C, 5% CO₂ humidified incubator.
- 9) Change Medium every 2 to 3 times a week.
- 10) Record each subculture event as a passage.

C. Maintenance and Generation of Seed Stocks

- 1) Change media the day after seeding and every 3-4 days thereafter. Use 100 mL of growth medium per T525 flask.
- 2) Following first or second passage after receipt of cells and with sufficient number of cells to continue maintenance and expansion, the major portion of the flasks should be sub-cultured using TrypLE as above under “Sub-culture” and a small portion should be set aside as a seed stock. The cell pellet for the seed stock should be resuspended in freezing medium.
- 3) Cells in freezing medium are dispensed into cryovials (1-2 million cells per 1mL aliquot) and frozen in a -80°C cryo-freezing container overnight.
- 4) Cryovials are transferred the next day to liquid nitrogen freezer for long-term storage.

D. Harvest

- 1) Passage cells until the desired number of cells is reached.
- 2) Remove cells from flasks as described above under “Sub-culture”.
- 3) Examine viability using Trypan blue staining.