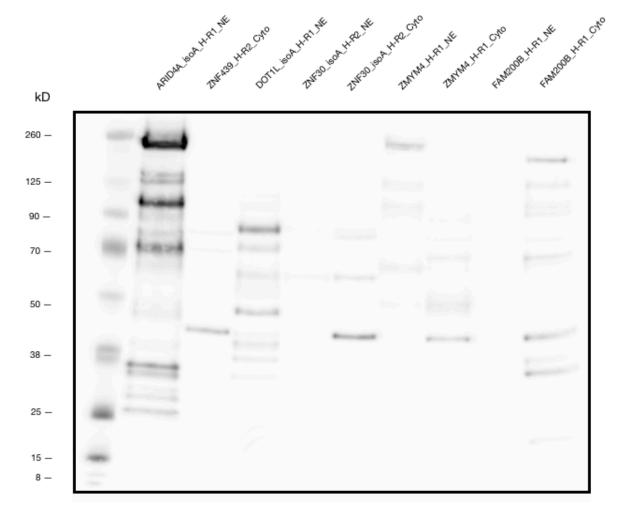
ARID4A (Homo sapiens), ZNF439 (Homo sapiens), DOT1L (Homo sapiens), ZNF30 (Homo sapiens), ZNF30 (Homo sapiens), ZMYM4 (Homo sapiens), FAM200B (Homo sapiens), and FAM200B (Homo sapiens)

Method:

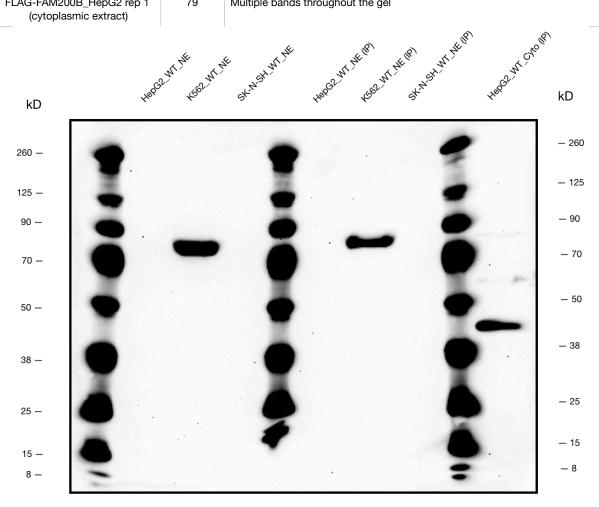
Western Blot Validation

Caption:

Each FLAG-tagged sample was immunoprecipitated from its corresponding protein isolate (500 uL - nuclear; 1 mL - cytoplasmic) using the FLAG Immunoprecipitation Kit (Sigma-Aldrich; cat# FLAGIPT1). The final elution step was performed by suspending the sample-bound resin in NuPage Sample Reducing Agent 10X and NuPage LDS Sample Buffer 4X (Thermo Fisher Scientific) and heating for 3 minutes at 90C. Followed by cooling on ice, the protein samples were loaded onto a NuPage 4-12% Bis-Tris gel (Thermo Fisher Scientific) and separated using a PowerEase 90W system (Thermo Fisher Scientific) running at 150 V for 1 hour. The protein bands were transferred to a nitrocellulose membrane using the Invitrogen iBlot 2 System (Thermo Fisher Scientific), and blocked overnight at 4C in 5% milk solution with gentle rocking. The membrane was treated with a 1:5000 dilution of monoclonal M2-Peroxidase-conjugated ANTI-FLAG antibody (diluted in 5% BSA solution) (Sigma-Aldrich; cat# A8592) for 1 hour. Following five 5-minute washes with 1X TBST, visualization was attained with the Super Signal West Femto solution kit (Thermo Fisher Scientific) and a MyECL Imager (Thermo Fisher Scientific). The second western blot image depicts negative control IPs prepared with HepG2 nuclear lysate (lane 6) and HepG2 cytoplasmic lysate (Lane 10).



| Lane | Loaded Sample | Expected Band Size (kDa) | Comments |
|------|---|--------------------------------|---|
| 1 | Ladder | N/A | N/A |
| 2 | FLAG-ARID4A_isoA_HepG2 rep 1 (nuclear extract) | 146 | Predicted size was 146 kDa. The observed size was 250 kDa, which is within 20% of an observer band of 220 kDa seen in https://www.novusbio.com/products/ retinoblastoma-binding-protein-1-antibody_nbp1-26616. The bands below may be degradation products. PTMs: Isopeptide bonding, Phosphorylation, and Ubl conjugation |
| 3 | FLAG-ZNF439_HepG2 rep 2 (cytoplasmic extract) | 61 | Single non-specific band seen in the negative control |
| 4 | FLAG-DOT1L_isoA_HepG2 rep 1 (nuclear extract) | 168 | Multiple bands below 20% of the expected size. PTMs: Phosphorylation |
| 5 | FLAG-ZNF30_isoA_HepG2 rep 2 (nuclear extract) | 74 | No visible banding |
| 6 | FLAG-ZNF30_isoA_HepG2 rep 2 (cytoplasmic extract) | 74 | Faint banding, with a darker non-specific band around 45 kDa |
| 7 | FLAG-ZMYM4_HepG2 rep 1 (nuclear extract) | 176 | Predicted size was 176 kDa. The observed size was 230 kDa, which is within 20% of an observed band of 220 kDa seen in https://www.novusbio.com/products/zmym4-antibody_nb100-94999 . PTMs: Acetylation, Isopeptide bonding, Phosphorylation, and Ubl conjugation |
| 8 | FLAG-ZMYM4_HepG2 rep 1 (cytoplasmic extract) | 176 | Banding smaller than the expected size. PTMs: Acetylation, Isopeptide bonding, Phosphorylation, and Ubl conjugation |
| 9 | FLAG-FAM200B_HepG2 rep 1 (nuclear extract) | 79 | No visible banding |
| 10 | FLAG-FAM200B_HepG2 rep 1 (cytoplasmic extract) | 79 | Multiple bands throughout the gel |



| Lane | Loaded Sample | Expected Band Size (kDa) | Comments |
|------|--|--------------------------------|---|
| 1 | Ladder | N/A | N/A |
| 2 | HepG2 Wild-Type (nuclear extract) | None | No visible banding |
| 3 | K562 Wild-Type (nuclear extract) | None | Single non-distinct band at around 80 kDa |
| 4 | SK-N-SH Wild-Type (nuclear extract) | None | No visible banding |
| 5 | Ladder | N/A | N/A |
| 6 | HepG2 Wild-Type (nuclear extract IP) | None | No visible banding |
| 7 | K562 Wild-Type (nuclear extract IP) | None | Single non-distinct band at around 80 kDa |
| 8 | SK-N-SH Wild-Type (nuclear extract) | None | No visible banding |
| 9 | Ladder | N/A | N/A |
| 10 | HepG2 Wild-Type (cytoplasmic extract IP) | None | Dark non-distinct band at around 45 kDa, and two lighter bands at about 60 kDa and 80 kDa |

Submitted by:Mark Mackiewicz and Michael Betti

Richard Myers, HAIB

Grant:

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